

Instruction for Use

【Product Name】

Carcinoembryonic Antigen (eCLIA)

【Order Information】

REF NO.	Package Size
0320500501	50 T
0320500502	2×50 T
0320500503	100 T
0320500504	2×100 T

【Intended Use】

Immunoassay for in vitro quantitative determination of Carcinoembryonic Antigen (CEA) in human serum and plasma.

Carcinoembryonic Antigen (CEA), a glycoprotein, was first identified by Gold and Freedman from fetal and colon cancer tissues in 1965. It is an embryonic carcinogenic antigen with a molecular weight of about 180KDa^{1,2}.

Serum CEA is a broad - spectrum tumor marker. Clinically, it can be used for auxiliary diagnosis of colon cancer, rectal cancer, lung cancer, breast cancer, esophageal cancer, pancreatic cancer, gastric cancer, metastatic liver cancer and other common tumors. Other malignant tumors such as medullary thyroid cancer, cholangiocarcinoma, urinary malignant tumors also have varying degrees of positive rate^{3,4}. In smokers, pregnancy, colitis, colonic polyps, intestinal diverticulitis, pancreatitis, liver cirrhosis, hepatitis, benign pulmonary diseases and cardiovascular diseases, serum CEA can also be increased to varying degrees, but the percentage of positive is low^{5,6}. Serum CEA level is one of the factors to judge the prognosis of tumor, and the continuous increase of serum CEA indicates poor prognosis. The serum CEA concentration of patients with elevated CEA before treatment decreased to normal level if surgery, chemotherapy, targeted therapy or immunotherapy were effective. If the serum CEA only partially or not decreased after treatment, the treatment effect is not good^{7,8}. Serum CEA can be used for follow-up and recurrence monitoring after tumor treatment. Generally, it should be tested every 2-3 months within 2 years after treatment and every 6 months within 3 to 5 years^{9,10}.

【Principles of the testing method】

Sandwich principle. Total duration of assay: 9 minutes.

The Carcinoembryonic Antigen (CEA) Assay Kit adopts the double-site sandwich electrochemiluminescence method. Antigen in the sample, biotinylated CEA monoclonal antibody and the ruthenium (Ru) complex-labeled CEA monoclonal antibody form an antigen-antibody sandwich complex, which is bound to streptavidin coated magnetic particles, and then transferred to the measuring cell where the magnetic particles are captured onto the surface of the electrode, while the unbound substances are washed away. Chemiluminescence is generated after the electrode is electrified, and the generated optical signal is measured by a photomultiplier, processed by an instrument, and the concentration of CEA in the sample is calculated based on the calibration curve.

【Main Components】

The reagent pack consists of MB, RA, RB, calibrators and control materials (optional), and different components and reagents of lot numbers should not be used interchangeably.

Components	Ingredients	Volume (2×50 T)	Volume (50 T)	Volume (100 T)	Volume (2×100 T)
(MB)	Streptavidin coated magnetic particles, 0.45 mg/mL; 0.1M phosphate-buffered saline (PBS); ProClin300	2×1.8 mL	1×1.8 mL	1×3.5 mL	2×3.5 mL
(RB)	Biotinylated CEA antibody, 1.5 mg/L; 0.1 M PBS; ProClin300	2×3.5 mL	1×3.5 mL	1×7.0 mL	2×7.0 mL
(RA)	Ru complex-labeled CEA antibody, 1.5 mg/L; 0.1 M PBS; ProClin300	2×3.0 mL	1×3.0 mL	1×6.0 mL	2×6.0 mL
Calibrator	CEA antigen, 0.05 M Tris buffer, ProClin 300	High: 2×1.0 mL Low: 2×1.0 mL	High: 1×1.0 mL Low: 1×1.0mL		
Control Material (Optional)	CEA antigen, 0.05 M Tris buffer, ProClin 300	High: 2×1.0 mL Low: 2×1.0 mL	High: 1×1.0 mL Low: 1×1.0 mL		

The assignment process of supporting calibrators in this pack is strictly implemented with reference to ISO 17511:2020, which can be traced to the WHO reference standard 73/601 of the first-generation IRP.

Refer to the quality control card for the target value and range of the quality controls.

Supporting instruments and materials required but not provided in this pack (provided by Lifotronic)

Additional materials for Automated ECL Immunoassay Analyzer eCL8000, eCL8000i, eCL8000p, eCL8000x:

- Auffer, 480 mL, 6×480 mL
- Buffer, 480 mL, 6×480 mL
- Concentrated Washing Buffer
- Assay Cup

Additional materials for Automated ECL Immunoassay Analyzer eCL9000, eCL9000i, eCL9600, eCL9900, eCL9900i:

- Auffer, 2 L
- Buffer, 2 L
- Concentrated Washing Buffer
- Assay Cup, reaction cup
- Disposable Sampling Head, pipette tips
- PreClean, system wash solution
- Waste Bags, litter bags

【Storage and Shelf Life】

Unopened reagent kit, calibrators and control materials should be placed at 2~8°C and will be valid for 18 months.

Once opened, they can be stored at 2~8°C for 56 days, and reagent kit can also be stored in machine (4~15°C).

The manufacturing date is labeled on the box, kit and bottles, and the expiration time is 18 months after production.

Damaged, expired or contaminated reagents should be discarded.

【Applicable Instrument】

Automated ECL Immunoassay Analyzer: eCL8000, eCL8000i, eCL8000p, eCL8000x, eCL9000, eCL9000i, eCL9600, eCL9900, eCL9900i.

【Specimen collection, preparation and storage】

It is recommended to use human serum samples or plasma samples collected with EDTA-2K, EDTA-3K, heparin lithium, heparin sodium blood collection tubes.

Blood samples should be collected in accordance with the standard operation of venipuncture. After the sample is completely coagulated, centrifugation should be performed to remove residual cellular substances. The sample should be free of air bubbles during testing. Lipid layer covering the sample after centrifugation should be removed. It is not recommended to use hemolyzed samples.

Samples can be stored for 7 days at room temperature (18~28°C), 14 days at 2~8°C, and 6 months at -15°C or below. Samples should avoid repeated freeze-thaw cycles. Do not use the samples after five freeze-thaw cycles.

【Testing Procedure】

Testing procedure and precautions

Before testing, the system operation manual of the measuring instrument should be carefully read, so as to obtain relevant information such as system operating procedures, sample management, safety precautions and maintenance, and materials required for testing should be prepared. CEA testing procedures should be called and set up in accordance with the system operating procedures.

Before using the reagent, put it into the analyzer 30 minutes in advance to automatically stir the magnetic bead particles and keep them in suspension.

Recommended environment temperature for testing: 10~30°C; relative humidity: ≤ 80%.

The CEA test adopts the double-site sandwich electrochemiluminescence method, with the total test time of 9 minutes, and the test method is as follows:

Step 1: After the user applies for testing, the system automatically aspirates 70 μL of biotinylated CEA monoclonal antibody, 60 μL of Ru complex-labeled CEA monoclonal antibody, 35 μL of streptavidin coated magnetic particles and 10 μL of sample into the reaction cup. They are automatically incubated at 37°C for 9 min to form antigen-antibody sandwich complexes, and then the whole complexes are bound to the magnetic particles under the interaction of biotin and streptavidin.

Step 2: After incubation, the system automatically aspirates the reaction mixture into the measuring cell, the magnetic particles are captured onto the surface of electrode, while the unbound substances are washed away by buffer, and the electrodes is applied with voltage to generate chemiluminescence. The generated optical signals are measured by the photomultiplier, and the measured results are automatically determined via the calibration curve specifically generated by the instrument (this curve is obtained by performing two-point calibration for the master calibration curve obtained by reading the reagent RFID).

Calibration

Calibration should be performed using the lot-matching reagent and calibrators.

The target value of the calibrator has been written into the Radio-Frequency Identification card (RFID card) of the kit.

1. Before calibration, the target value, reagent information and master calibration curve information of the calibrator in the RFID card (Radio Frequency Identification, inductive electronic chip or proximity card) of the kit should be imported into the test system by swiping the card.
2. The supporting calibrator is tested, and the analyzer adjusts the master calibration curve according to the test results of the supporting calibrator to obtain the calibration curve tested by the current system (the analyzer can automatically interpret the validity of the calibration curve according to the adjustment results).

The test system should re-perform the calibration operation under the following

conditions:

- (1) When different lots of reagents are used;
- (2) When the same lot of reagents has been used on the analyzer for more than 28 days;
- (3) According to requirements: e.g. if the quality control result exceeds the defined limit;
- (4) When changing the buffers of different lot numbers.

Quality Control procedure

In order to ensure the reliability of test results, it is recommended to test the control materials every 24 hours. It is recommended to test the control materials after each calibration, reagent lot replacement, maintenance or troubleshooting. The quality control results should all fall within the defined range. If they exceed the defined range, the reasons such as instrument status, reagents and calibrators should be investigated and the corrective measures should be taken.

Calculation

The system software can automatically calculate the analyte concentration, with the result in ng/mL or µg/L.

1 ng/mL CEA is equivalent to 16.9 mIU/mL.

Specimen dilution

CEA concentration in the sample higher than the upper limit of detection can be diluted with sample diluent.

The recommended dilution ratio is 1:20 (automatic dilution by instrument or manual dilution). If manual dilution, the result should be multiplied by the dilution ratio. If instrument automatic dilution, the instrument will automatically calculate the result.

Biological Reference Interval

By analyzing the serum samples of healthy people, it was calculated that the upper limit of the reference range of 95% is as follows.

Analysis on reference interval	All subjects	Non-smokers	Smokers
Age (years)	18-70	18-70	18-70
95 percentile	4.702	3.801	5.515
Quantity	241	121	120

Due to the differences in geography, race, gender and age, it is recommended that each laboratory determine the applicability of reference range through tests, and establish the reference range of this laboratory if necessary.

Interpretation of results

When interpreting the test results, it is necessary to refer to the patient's overall clinical situation, including: symptoms, medical history as well as other corresponding data and information.

Limitations

The test results are only for clinical reference and cannot be used alone as the basis for diagnosis or exclusion of cases.

The detection range of the kit is 0.1~1000 ng/mL. If the CEA concentration in the sample is lower than the lower limit of detection, the result is reported as < 0.1 ng/mL; if the CEA concentration in the sample is higher than the upper limit of detection, the result is reported as > 1000 ng/mL (20x diluted sample is reported as > 20000 ng/mL).

When jaundice (bilirubin) is ≤ 66 mg/dL, hemolysis (hemoglobin) is ≤ 1100 mg/dL, lipemia (triglyceride) is ≤ 1500 mg/dL, biotin is ≤ 20 ng/mL, total protein is ≤ 10 g/dL, and HAMA is ≤ 100 ng/mL in the sample, the interference deviation of the measured results is within ±10.0%. When the concentration of rheumatoid factor in the sample is ≤ 1500 IU/mL, the relative recovery of the test results is between 90% and 110%.

When the sample contains adriamycin (100 µg/mL), 5-fluorouracil (360 µg/mL), methotrexate (4500 µg/mL), cisplatin (1.5 µg/mL) and cyclophosphamide (3000 µg/mL), the interference deviation of the measured results is within ±10.0%.

Samples of other tumor markers containing 1000 ng/mL AFP, 1000 U/mL CA125, 100 U/mL CA15-3, 1000 U/mL CA19-9, 100 ng/mL PSA and 1000 ng/mL Ferr are tested, and the CEA measured results are not greater than 0.2 ng/mL.

When the CEA concentration reaches 10000 ng/mL, there is no high-dose hook effect; when the CEA concentration reaches 200000 ng/mL, the test results are not affected by the hook effect.

Analytical Performance

Lower limit of measurement

Limit of Detection (LoD) ≤ 0.2 ng/mL.

Accuracy

The relative deviation of its measured results is within ±10.0%.

Linearity

Within the range of 0.2 ng/mL~1000 ng/mL, the correlation coefficient (r) of the kit should not be less than 0.9900.

Within-run imprecision (repeatability)

The coefficient of variation (CV) ≤ 5.0%.

Between-lot imprecision

The coefficient of variation (CV) ≤ 10.0%.

Homogeneity of calibrators and control materials

In-bottle homogeneity: coefficient of variation (CV) ≤ 5.0%.

Accuracy of calibrators

The relative deviation of its measured results is within ±10.0%.

Measured values of quality control

The measured results of supporting assigned quality control of the kit should be within the quality control range.

Precaution and Warning

This kit is only used for in vitro diagnosis;















When using this kit, the relevant operation precautions of the laboratory must be observed;

The test results of this kit can only be used as clinical reference, the patient's clinical evaluation should be based on the patient's clinical symptoms/signs, medical history, other laboratory test results and treatment response, etc.;

Due to methodological reasons or antibody specificity, the results of the same sample tested with reagents of different manufacturers may be different. Therefore, results obtained with different kits should not be directly compared, so as to prevent wrong medical interpretation; it is recommended that the Laboratory Department indicate the characteristics of the reagents in the test report issued to the clinician. If the reagent type is changed during series monitoring, continuous testing should be conducted, and the results should be compared with the original reagent results in parallel to re-determine the baseline value;

This product contains animal-derived substances, and may have potential biological risks. All samples and reaction wastes should be treated as the source of infection, and all wastes must be disposed of in accordance with local regulations.

Symbol

Symbol	Title of Symbol	Symbol	Title of Symbol
	Manufacturer		Consult instructions for use
	Use by		In vitro diagnostic medical device
	Lot number		Sufficient for <n> tests
	Serial number		Biological risks
	Temperature limitation		This way up
	Authorized representative in the European Community		Indicates this device is in compliance with Europe Directive.
	Catalogue number		Material code

References

1. Gold P, Freedman SO. Demonstration of tumor-specific antigens in human colonic carcinomata by immunological tolerance and absorption techniques. J Exp Med, 1965, 121: 439-462.
2. Gold P, Freedman SO. Specific carcinoembryonic antigens of the human digestive system. J Exp Med, 1965, 122: 467-481.
3. Go VLW, Zamcheck N. The role of tumor markers in the management of colorectal cancer. Cancer, 1982;50:2618-2623.
4. Sell SS. Serological cancer markers. Humana Press, 1992; ISBN 0-89603-209-4.
5. Fukuda I, Yamakado M, Kiyose H. Influence of Smoking on Serum Carcinoembryonic Antigen Levels in Subjects Who Underwent Multiphasic Health Testing and Services. J Med Syst 1998;22(2): 89-93.
6. Ruibal Morell A. CEA serum levels in nonneoplastic disease. Int J Biol Markers 1992;7(3):160-166.
7. von Kleist S, Chavanel G, Burtin P. Identification of an antigen from normal human tissue that cross-reacts with the carcinoembryonic antigen. Proc Natl Acad Sci USA, 1972;69:2492-2494.
8. Matsuoka Y, Hara M, Takatsu K, Kitagawa M. Presence of antigen related to the carcinoembryonic antigen in feces of normal adults. GANN, 1973;64: 203-206.
9. Duffy MJ, Van Dalen A, Haglund C, et al. Clinical utility of biochemical markers in colorectal cancer: European Group on Tumour Markers (EGTM) guidelines. Eer J Cancer 2003;39(6):718-727.
10. Sturgeon CM, Duffy MJ, Stenman UH, et al. National Academy of Clinical Biochemistry Laboratory Medicine Practice Guidelines for Use of Tumor Markers in Testicular, Prostate, Colorectal, Breast and Ovarian Cancers. Clin Chem, 2008;54(12):e11-e79.

Manufacturer

Shenzhen Lifotronic Technology Co., Ltd.
Unit A, 4th Floor, Building 15, Yijing Estate, No.1008 Songbai Road, Nanshan District, Shenzhen City, Guangdong Province, 518055, P.R.China
E-mail: inter-service@lifotronic.com

European Representative

Shanghai International Holding Corp. GmbH (Europe)
Eiffestrasse 80, 20537 Hamburg, Germany
Tel: +49-40-2513175 Fax: +49-40-255726

【Version and Revision】

Version: A0

Issue Date: 2021-08-01